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## 54) Title: A COLLAGEN BINDING PROTEIN AS WELL AS ITS PREPARATION

## (57) Abstract

The present invention relates to a new recombinant hybrid-DNA-molecule comprising a nucleotide sequence from *S. aureus* coding for a protein, or polypeptide, having collagen binding properties.

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A COLLAGEN BINDING PROTEIN AS WELL AS ITS PREPARATION.

DESCRIPTION

Technical field

The present invention relates to a collagen binding protein as well as hybrid-DNA-molecules, e.g. plasmids or phages comprising a nucleotide sequence coding for said protein. Further the invention relates to micro-organisms comprising said molecules and their use producing said protein, as well as the synthetic preparation of said protein. In particular the invention relates to a cloned gene encoding the *Staphylococcus aureus* collagen binding protein, or functionally active portions thereof, vectors containing the cloned gene or parts thereof, and micro-organisms transformed by those vectors as well as the cloning of the gene which specify the biosynthesis of *Staphylococcus aureus* collagen binding protein (CBP) (also called the collagen receptor by Switalski et al 1989) and the use of organisms transformed with the cloned gene to produce CBP or CBP like proteins. The invention also describes the use of this gene for diagnostic purposes.

The object of the present invention is to obtain a collagen binding protein.

A further object is to obtain said protein by means of a genetic engineering technique by using e.g. a plasmid comprising a nucleotide sequence coding for said protein.

A further object is to obtain a possibility of preparing said protein by chemical synthesis.

Further objects will be apparent from the following description.

Background of the invention

WO-A1-85/05553 discloses bacterial cell surface proteins having fibronectin, fibrinogen, collagen, and/or laminin binding ability. Thereby it is shown that different bacteria have an ability to bind to fibronectin, fibrinogen, collagen, and/or laminin.

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TACAAAACCA AAATTACGAA TGAACAGCAA AA<sup>2</sup>GAGTTG TTAATAATTC  
ACAAGCTGG TATCAAGAGC ATGGTAAGGA AC AGTGAAC GGGAAATCAT  
TTAACATAC TGTGCACAAT ATTAATGCTA ATGCCGGTAT TGAAGGTACT  
GTAAAAGGTG AATTAAAAGT TTTAAAACAG GATAAAGATA CCAAGGCTCC  
5 TATAGCTAAT GTAAAATTAA AACTTTCTAA AAAAGATGGA TCAGTTGTAA  
AGGACAATCA AAAAGAAATT GAGATTATAA CAGATGCAA CGGTATTGCT  
AATATTAAAG CGTTGCCTAG TGGAGACTAT ATTTAAAAG AAATAGAGGC  
GCCACGACCG TATACATTG ATAAGGATAA AGAATATCCG TTTACTATGA  
AAGATACAGA TAATCAGGGA TATTTACGA CTATTGAAAA TGCAAAAGCG  
10 ATAGAAAAAA CAAAAGATGT TTCTGCTCAA AAGGTTGGG AAGGCACTCA  
AAAAGTAAA CCAACGATTT ATTCAGTT GTACAAACAA GATGACAATC  
AAAATACAAC ACCAGTAGAC AAAGCAGAGA TTAAAAAATT AGAAGATGGA  
ACGACAAAAG TGACATGGTC TAATCTTCCG GAAAATGACA AAAATGGCAA  
GGCTATTAAA TATTTAGTTA AAGAAGTAAA TGCTCAAGGT GAAGATAACAA  
15 CACCAGAAGG ATATACTAAA AAAGAAAATG GTTAGTGGT TACTAATACT  
GAAAAACCAA TCGAAACAAAC ATCAATTAGT GGTGAAAAAG TATGGGACGA  
CAAAGACAAT CAAGATGGTA AGAGACCAGA AAAAGTCAGT GTGAATTTAT  
TGGCTAACCG GGAGAAAGTA AAAACGTTAG ACGTGACATC TGAAACAAAC  
TGGAAAGTACG AATTAAAGA CTTACCGAAG TATGATGAAG GAAAGAAAAT  
20 AGAATATACA GTGACCGAAG ATCACGTAAA AGACTACACA ACAGACATCA  
ACGGTACGAC AATAACGAAC AAGTATACAC CAGGAGAGAC ATCGGCAACA  
GTAACAAAAA ATTGGGATGA CAATAATAAC CAAGACGGAA AACGACCAAC  
TGAAATCAA GTTGAGTTAT ATCAAGACGG AAAAGCAACA GGAAAAACGG  
CAACATTAAA TGAATCTAAT AACTGGACCC ATACGTGGAC AGGATTAGAT  
25 GAAAAAGCAA AAGGACAACA AGTAAAATAC ACAGTCGAGG AATTAACAAA  
GGTCAAAGGT TATACAACAC ATGTGGATAA CAATGATATG GGTAACTTGA  
TTGTGACGAA TAAATATACG CCAGAAACAA CATCAATTAG TGGTAAAAAA  
GTATGGACG ACAAAAGACAA TCAAGATGGT AAGAGACCAG AAAAAGTCAG  
TGTGAATTTA TTGGCTGATG GAGAGAAAGT AAAAACGTTA GACGTGACAT  
30 CTGAAACAAA CTGGAAGTAC GAATTTAAAG ACTTACCGAA GTATGATGAA  
GGAAAGAAAA TAGAATATAC AGTGACCGAA GATCACGTAA AAGACTACAC  
AACAGACATC AACGGTACGA CAATAACGAA CAAGTATACA CCAGGAGAGA  
CATCGGCAAC AGTAACAAAA AATTGGGATG ACAATAATAA CCAAGACGGA  
AAACGACCAA CTGAAATCAA AGTTGAGTTA TATCAAGACG GAAAGCAAC  
35 AGGAAAAACG GCAACATTAA ATGAATCTAA TAACTGGACC CATACTGGG  
CAGGATTAGA TGAAAAGCA AAAGGACAAC AAGTAAAATA CACAGTCGAG

- 5 -

TyrGlySerTyrAsnSerPheSerIleAsnTyrLysThrLysIleThrAsnGluGlnGln  
LysGluPheValAsnAsnSerGlnAlaTrpTyrGlnGluHisGlyLysGluGluValAsn  
GlyLysSerPheAsnHisThrValHisAsnIleAsnAlaAsnAlaGlyIleGluGlyThr  
ValLysGlyGluLeuLysValLeuLysGlnAspLysAspThrLysAlaProIleAlaAsn  
5 ValLysPheLysLeuSerLysLysAspGlySerValValLysAspAsnGlnLysGluIle  
GluIleIleThrAspAlaAsnGlyIleAlaAsnIleLysAlaLeuProSerGlyAspTyr  
IleLeuLysGluIleGluAlaProArgProTyrThrPheAspLysAspLysGluTyrPro  
PheThrMetLysAspThrAspAsnGlnGlyTyrPheThrThrIleGluAsnAlaLysAla  
IleGluLysThrLysAspValSerAlaGlnLysValTrpGluGlyThrGlnLysValLys  
10 ProThrIleTyrPheLysLeuTyrLysGlnAspAspAsnGlnAsnThrThrProValAsp  
LysAlaGluIleLysLysLeuGluAspGlyThrThrLysValThrTrpSerAsnLeuPro  
GluAsnAspLysAsnGlyLysAlaIleLysTyrLeuValLysGluValAsnAlaGlnGly  
GluAspThrThrProGluGlyTyrThrLysLysGluAsnGlyLeuValValThrAsnThr  
GluLysProIleGluThrThrSerIleSerGlyGluLysValTrpAspAspLysAspAsn  
15 GlnAspGlyLysArgProGluLysValSerValAsnLeuLeuAlaAsnGlyGluLysVal  
LysThrLeuAspValThrSerGluThrAsnTrpLysTyrGluPheLysAspLeuProLys  
TyrAspGluGlyLysLysIleGluTyrThrValThrGluAspHisValLysAspTyrThr  
ThrAspIleAsnGlyThrIleThrAsnLysTyrThrProGlyGluThrSerAlaThr  
ValThrLysAsnTrpAspAspAsnAsnAsnGlnAspGlyLysArgProThrGluIleLys  
20 ValGluLeuTyrGlnAspGlyLysAlaThrGlyLysThrAlaThrLeuAsnGluSerAsn  
AsnTrpThrHisThrTrpThrGlyLeuAspGluLysAlaLysGlyGlnGlnValLysTyr  
ThrValGluGluLeuThrLysValLysGlyTyrThrThrHisValAspAsnAsnAspMet  
GlyAsnLeuIleValThrAsnLysTyrThrProGluThrThrSerIleSerGlyGluLys  
ValTrpAspAspLysAspAsnGlnAspGlyLysArgProGluLysValSerValAsnLeu  
25 LeuAlaAspGlyGluLysValLysThrLeuAspValThrSerGluThrAsnTrpLysTyr  
GluPheLysAspLeuProLysTyrAspGluGlyLysLysIleGluTyrThrValThrGlu  
AspHisValLysAspTyrThrThrAspIleAsnGlyThrThrIleThrAsnLysTyrThr  
ProGlyGluThrSerAlaThrValThrLysAsnTrpAspAspAsnAsnAsnGlnAspGly  
LysArgProThrGluIleLysValGluLeuTyrGlnAspGlyLysAlaThrGlyLysThr  
30 AlaThrLeuAsnGluSerAsnAsnTrpThrHisThrTrpThrGlyLeuAspGluLysAla  
LysGlyGlnGlnValLysTyrThrValGluGluLeuThrLysValLysGlyTyrThrThr  
HisValAspAsnAsnAspMetGlyAsnLeuIleValThrAsnLysTyrThrProGluThr  
ThrSerIleSerGlyGluLysValTrpAspAspLysAspAsnGlnAspGlyLysArgPro  
GluLysValSerValAsnLeuLeuAlaAsnGlyGluLysValLysThrLeuAspValThr  
35 SerGluThrAsnTrpLysTyrGluPheLysAspLeuProLysTyrAspGluGlyLysLys  
IleGluTyrThrValThrGluAspHisValLysAspTyrThrThrAspIleAsnGlyThr

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The invention further comprises a microorganism containing at least one hybrid-DNA-molecule according to the above. The plasmid pSAC104 in an E. coli strain TG1 has been deposited at the Deutsche Sammlung von Mikroorganismen (DSM), and has thereby been allocated the deposition number DSM 6199. The present invention provides a cloned gene encoding the CBP having improved CBP-properties as compared with native CBP which is released and purified from *S. aureus* cells. The gene is derived from a *S. aureus* strain and inserted into a cloning vector. Cells of a prokaryotic organism which have been transformed with recombinant vectors are disclosed.

The invention further provides the identification of the nucleotide sequence of the gene encoding the CBP here called the cbp-gene. The deduced amino acid sequence reveals a molecule with several distinct features resembling staphylococcal cell surface proteins.

The invention also provides a procedure for production and purification of the recombinant CBP. This is done in a way so that the molecule retains its collagen binding properties, thus this recombinant CBP resembles the native unreleased *S. aureus* CBP.

The invention further provides the use of the cbp-gene for diagnostic purposes. Gene probes chosen to specifically recognize the presence of the cbp gene in clinical *S. aureus* isolates have been used. As an example, the correlation between the presence of CBP on the surface of *S. aureus* strains isolated from patient with septic arthritis could be verified by the presence of the cbp-gene in all tested strains.

Appropriate carrier proteins can be coupled to the amino acid sequence as well, such as IgG binding regions of protein A.

The invention will be described in the following with reference to the examples given, however, without being restricted thereto.

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S. aureus fibronectin receptor (●, ZZFR). Panel A - binding of  $^{125}\text{I}$ -collagen to protein coated beads as a function of time. Panel B - inhibition of binding of  $^{125}\text{I}$ -collagen by antibodies. Attachment of  $^{125}\text{I}$ -labeled beads to cartilage as a function of time (panel C) and inhibition of attachment of  $^{125}\text{I}$ -labeled beads to cartilage by antibodies (panel D). In this experiment 1 ug of adhesin protein was coupled to  $10^8$  polystyrene beads. Control beads were coated with the same molar concentration of the fibronectin receptor. Unreacted sites on the beads were saturated with bovine serum albumin. Scanning electron microscopy of beads coated with collagen adhesin (panel E) or fibronectin receptor protein (panel F) attached to cartilage.

Figure 6: Expression constructs utilized to localize the collagen binding domain within the S. aureus collagen adhesin.

Example 1:

20 Cloning and identification of the cbp-gene in E.coli

In order to isolate the gene encoding S. aureus CBP two commercial available (Clontech laboratories, Inc. Palo Alto, CA, USA) S. aureus strains (strain FDA 574 and FDA 485) were tested if they bound radioactivity labelled 25 collagen. This was done according to Switalski et al 1989. Strain 574 was found to bind collagen and therefore a gene library (obtained from the same company, cat. #XL 15016) of the same strain was screened for the expression of CBP activity. Using the suppliers protocoll (in addition to 30 this protocoll the general work involving molecular genetic appropriate protocols found in "Current Protocols in Molecular Biology" Vol. 1 and 2, (edited by Ausubel, F.M., R. Brent, R.L. Kingston, D.D. Moore, I.G. Seidman, J.A. Smith, U. Struhl, Greene, Wiley Interscience), and 35 "Molecular Cloning". A laboratory manual, (Maniatis, T., Fritsch, E.F. and J. Sambrook (1982) Cold Spring Harbor

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to detect bound primary Fab-Fragments. After incubation for 1 h at RT the filters were washed 3 X 10 min in PBS-T. The bound labelled secondary antibodies were detected by a color reaction according to the manufacturer's instructions 5 (Bio-Rad, Instructions for preparing the BCIP/NBT color development solution for use in the immun-blot alkaline phosphatase assay kit).

By the use the above described methods several recombinant lambda phages expressing CBP-activity could be 10 identified and isolated.

Two of these were chosen for further studies. They were called lambda coll 1 and lambda cCOLR6A respectively.

Subcloning lambda coll 1: Purified lambda coll 1 15 DNA was cleaved with EcoRI and the sticky ends were filled in using Klenow fragments together with the dNTP's. The blunt ended DNA-fragments originating from the *S. aureus* chromosome were ligated into Sma 1 cleaved pUC 18 (Pharmacia-LKB Biotechnology, Uppsala, Sweden). After transformation into freeze competent *E. coli* TG1 cells recombinant 20 clones were tested for expression of the CBP. It was found that all clones expressing CBP harboured a recombinant plasmid with an insert of approx. 4 kb. One such clone called p 16 was chosen for further studies and a schematic 25 map of the insert in this clone is shown in Fig. 1 A.

In a similar way as lambda coll 1 two other lambda clones were generated from the screening of the genomic library. Large scale cultures of pure positives were obtained and the DNA was isolated. EcoRI digestion of the 30 clones resulted in inserts with two different sizes. Clone 1A had an insert of 3.2 kb and 3B had an insert of 4.5 kb. The larger of the two was used for further characterization. Purified insert DNA (1.5 kb) from  $\lambda$  GT11 clone 3B was 35 ligated to EcoRI digested puc18 and transformed into *E. coli* TB-1 cell creating subclone cCOLR6A. It was also subcloned into M13mp18/JM101 for sequencing.

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structure found in staphylococcal protein A (Guss et al 1984) and FnBP A (Signäs et al 1989) as well as streptococcal protein G (Guss et al 1986) and M protein (Hollingshead et al 1986). This region is thought to mediate the 5 binding of the protein to the cell wall. The amino acid sequence nearest to the C-terminal end consists of a long stretch of hydrophobic residues followed by some charged amino acids. This region called M is similar in structure to the C-terminal end of protein A, FnBP A, Protein G and M 10 protein.

The predicted mol.wt of the deduced CBP is approx. 133 kd (including the postulated signal sequence, S) which is very close to the mol.wt of 135 kd reported for the native released receptor (Switalski et al 1989).

15 In order to construct a plasmid coding for the complete cbp-gene *S. aureus* FDA 574 chromosomal DNA was purified and double cleaved with Hind III/Pst 1. With the guidance of Southern Transfer experiments using a 32-P labelled oligonucleotide probe (5'-ATTAAAGCGTTGCCTAGTGG-3') 20 it was known that cleavage with these enzymes should generate an approx. 3,2 kb fragment corresponding to the 3'end of the cbp-gene. After cleavage with these enzymes the chromosomal DNA was electrophoretically separated in an agarose gel. A gel slice ruffly corresponding to right size 25 was cut out and the DNA fragments eluted and purified. The purified fragments were ligated into pUC 18 previously double cleaved with Hind III/Pst 1. After ligation followed transformation into *E. coli* TG1 and the resulting recombinant clones were screened for obtaining the right fragment 30 using colony hybridization with the same probe. One positive clone hybridizing with the radioactive probe was chosen for further studies. This clone called *E. coli* pSAC 100 was cleaved with Hind III and a purified approx. 1,8 kb Hind III fragment from p 16 (encoding the 5'end of the cbp-gene, 35 Fig. 1 A) was ligated into pSAC 100. After transformation

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activity. This result is in contrast with the findings reported by Switalski et al 1989 which found that purified or partly purified native collagen receptor could not inhibit the binding of collagen to *S. aureus* Cowan 1 cells. The 5 conclusion of this is that recombinant CBP expressed has retained more of its original features than the released protein from the staphylococci.

Although it was possible to detect CBP activity in the recombinant *E. coli* lysate it was not possible to 10 affinity purify the CBP using immobilized collagen or gelatine. Although in "Western transfer" experiments with lysates from the above mentioned recombinant clones, using the Fab-fragments described in Example 1, was it possible to detect bands corresponding to high mol.wt. fragments. These 15 were in the same size as expected from calculations using the deduced amino acid sequence.

Example 4:

Expression and of a CBP fusionprotein which retains the  
20 collagen binding properties after purification

Been unsuccessful to affinity purify the recombinant produced CBP, using immobilized collagen, another approach was used. This approach was to fuse the cbp-gene or parts of the gene to another gene encoding a so called 25 affinity tail (Methods in enzymology, Part 185). The affinity tail to be tested was the part from the protein A gene encoding the IgG-binding domains (Uhlein et al 1984). Therefore a vector encoding the above mentioned domains from protein A was used. This vector called pNSEQ1, which 30 was a gift from Dr. M. Uhlein contains in addition to the IgG-binding domains (E, D, A, B and C) two multi cloning sites (MCS) which flank the IgG-binding domains. This makes it possible to chose a restriction enzyme that has a 35 recognition site in both the MCS which upon cleavage results in a release of (provided the restriction site is not present in the IgG-binding domains) a DNA fragment encoding the

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Example 5:

The use of the CBP-gene as a diagnostic tool

Two oligonucleotides (JP-1, 5'-AGT-GGT-TAC-TAA-TAC-TG-3' and JP-2, 5'-CAG-GAT-AGA-TIG-GTT-TA-3') complementary 5 to regions of the CBP that flanked the repeats B1, B2, and B3 were constructed (Oligo's Etc.). Genomic DNA from 6 different *Staphylococcus aureus* strains that were known to bind  $^{125}\text{I}$ -collagen (Table 1) were isolated as previously described by Lindberg. Polymerase chain reaction 10 (PCR) was performed with a Cetus/Perkin-Elmer DNA Thermo-cycler. Reaction mixtures (100  $\mu\text{l}$ ) contained 1mM of each primer, 200 mM of each dNTP, 1 mM Tris-HCl (pH 8.3), 5 mM KCl, 15 mM MgCl<sub>2</sub>, 0.001% gelatin, 3  $\mu\text{g}$  template DNA, and 2.5 U AmpliTaq DNA polymerase. The reaction mixtures were 15 overlayed with 100  $\mu\text{l}$  of mineral oil and amplified for 30 cycles consisting of a 2 minute denaturation at 94°C, a 2 minute annealing period at 55°C, and a 3 minute extension period at 72°C. After amplification, 15  $\mu\text{l}$  of the PCR products were analyzed on a 1% agarose gel (SeaKem GTG, FMC 20 Inc., Rockland, Maine).

PCR analysis of the genomic DNA from the different *S. aureus* isolates revealed two distinctly different sized products. FDA 574, Cowan, and #13 all had gene products of 1677 bp, whereas Phillips, #7, and #14391 had gene products 25 of 1118 bp. *S. aureus* Newman, a known non-collagen binder had no detectable PCR product. There is a direct correlation between the repeat size and the estimated molecular weight of the purified native collagen receptor from the different *S. aureus* strains tested. Upon further sequence analysis, it 30 appears that a PCR product of 1677 bp corresponds to 3 repeat units, each 560 bp long. A PCR product of 1118 bp therefore corresponds to 2 repeats, each 560 bp long. These data correlate highly with the estimated molecular weight of purified native collagen receptors of 135 kd and 115 kd 35 respectively.

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lysostaphin lysates prepared from different *S. aureus* isolates (Figure 3). Lysostaphin digestion releases from the cell surface of *S. aureus* a number of proteins, around 30 bands can be visualized in the lysates by Coomassie  
5 Brilliant Blue staining of the gel (Switalski et al., 1989). The anti-adhesin antibodies recognized a component of  $M_r$  135 kd in the lysate of strain Cowan (Figure 3, lane a), which is in agreement with our previous observations (Switalski et al., 1989). The major immunoreactive protein  
10 detected in the lysates of the other collagen adhesin positive strains (CA+) varied in molecular weight and was present as either 110 kd or 135 kd (Figure 3, lanes b through h). No correlation was observed between the apparent size of the immunoreactive protein and the collagen binding capacity  
15 of a strain or its origin (bone, synovial fluid). None of the nine non-binding collagen *S. aureus* strains tested expressed an immunoreactive protein (Figure 3, lane i).

20 Collagen adhesin mediated attachment of staphylococci to collagenous substrata.

The relationship between the ability to express a collagen adhesin and the observed localization of an infection within collagen rich tissues prompted us to analyze the role of the cell surface adhesin in bacterial attachment to  
25 collagen containing substrates. We initially studied attachment of bacteria to surfaces coated with type II collagen. Results indicated that a collagen coated surface was an excellent attachment substrate for strains which express a surface localized collagen adhesin. The attachment is time  
30 dependent and saturable reaching an equilibrium after 3 hours of incubation (Figure 4A). The number of attaching bacteria is not influenced by the size of the adhesin since strains #14 and Phillips, which either express a 135 kd or 110 kd adhesin respectively, attached in equal numbers to  
35 the collagen coated substrate. When bacteria were preincubated with anti-adhesin antibodies, against the collagen

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Creation of artificial bacteria

"Artificial bacteria" were prepared by covalently coating polystyrene beads (1.2  $\mu\text{m}$  vs. staphylococci 0.8 - 1.0  $\mu\text{m}$  in diameter) with the collagen adhesin protein. These 5 beads were then tested in a series of experiments analogous to those performed with intact bacteria. The collagen adhesin (CA) coated beads, but not beads coated with a recombinant form of another staphylococcal cell surface component, the fibronectin receptor (Flock et al., 1987), 10 bound  $^{125}\text{I}$ -collagen (Figure 5A) in a manner similar to that of CA+ strains of *S. aureus* (Speziale et al., 1986). This binding was abolished by anti-CA antibodies, whereas preimmune antibodies did not effectively inhibit binding 15 (Figure 5B). When "artificial bacteria" were assayed for the ability to attach to collagen (data not shown) or cartilage, we found that CA beads adhered to the substrate in a time dependent manner, identical to that of CA+ strains of *S. aureus*, while beads coated with the fibronectin receptor did not adhere at significant levels (Figure 5C). The anti-CA 20 antibody inhibited the adhesion of CA beads to cartilage in a dose dependent fashion, whereas a preimmune antibodies had no effect (Figure 5D). Once again the quantitative binding data was corroborated by electron microscopy observations. CA coated beads attached in large numbers to cartilage 25 tissue, in particular to collagen fibers (Figure 5E), while beads coated with the fibronectin receptor did not (Figure 5F).

Localization of the collagen binding domain within the 30 collagen adhesin.

Various expression constructs have been created in *E. coli* in effort to specifically localize the collagen binding domain. Two different types of expression vectors have been utilized in these experiments, pKK223-3 and 35 pGEX-2T, the second of which results in the collagen adhesin fused to glutathione-S-transferase. To date the smallest

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including man, the protein, or polypeptide is dispersed in sterile, isotonic saline solution, optionally while adding a pharmaceutically acceptable dispersing agent. Different types of adjuvants can further be used in order to sustain 5 the release in the tissue, and thus expose the protein or the peptide for a longer time to the immundefense system of a body.

A suitable dosage to obtain immunization is 0,5 to 5  $\mu$ g of CBP, or polypeptide, per kg bodyweight and 10 injection of immunization. In order to obtain a durable immunization, vaccination should be carried out at more than one consecutive occasion with an interval of 1 to 3 weeks, preferably at three occasions.

When using the present CBP, or polypeptide, for 15 topical, local administration the protein is dispersed in an isotonic saline solution to a concentration of 25 to 250  $\mu$ g per ml. The wounds are then treated with such an amount only to obtain a complete wetting of the wound surface. For an average wound thus only a couple of millilitres 20 of solution are used in this way. After treatment using the protein solution the wounds are suitably washed with isotonic saline or another suitable wound treatment solution.

Further the collagen binding protein as well as the minimal collagen binding site polypeptide, of the present 25 invention can be used to diagnose bacterial infections caused by Staphylococci strains, whereby a collagen binding protein of the present invention is immobilized on a solid carrier, such as small latex or Sepharose<sup>®</sup> beads, whereupon sera containing antibodies are allowed to pass and 30 react with the CBP thus immobilized. The agglutination is then measured by known methods.

Further, the CBP, or the polypeptide can be used in an ELISA test (Enzyme Linked Immuno Sorbent Assay; E Engvall, Med. Biol. 55, 193, (1977). Hereby wells in a 35 polystyrene microtitre plate are coated with the CBP, and incubated over night at 4°C. The plates are then thorough-

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References:

Carret, G., H. Emonard, G. Fardel, M. Druguet, D. Herbage, and J. P. Flandrois. 1985. Ann. Inst Pasteur (Paris) 5 136A:241-245.

Guss, B., M. Uhle'n, B. Nilsson, M. Lindberg, J. Sjöquist and J. Sjödahl. 1981. J. Biochem., 138, 413-420.

Guss, B., M. Eliasson, A. Olsson, M. Uhle'n, A.-K. Frej, H. Jörnvall, J.-I. Flock and M. Lindberg. 1986. EMBO J., 10 5, 1567-1575.

Holderbaum, D., R.A. Spech and L. A. Ehrhart. 1985. Collagen Relat. Res. 5:261-271.

Holderbaum, D., G. S. Hall and L. A. Ehrhart. 1986. Infect. Immun. 54:359-364.

15 Hollingshead, S. K., V. A. Fischetti and J. R. Scott. 1986. J. Biol. Chem. 261:1677-1686.

Signäs, S., G. Raucci, K. Jönsson, P.-E. Lindgren, G. M. Anantharamaiah, M. Höök and M. Lindberg. 1989. Proc. Nutl. Acad. Sci. USA. 86:699-703.

20 Speziale, P. G. Raucci, L. Visal, L. M. Switalski, R. Timpl and M. Höök. 1986. J. Bact. 167:77-81.

Switalski, L. M., P. Speziale and M. Höök. 1989. J. Biol. Chem. 264:21080-21086.

Uhle'n, M., B. Guss, B. Nilsson, S. Gatenbeck, L. Philipsson 25 and M. Lindberg. 1984. J. Biol. Chem. 259:1695-1702.

Vercellotti, G. M., J. B. McCarthy, P. Lindholm, P. K. Peterson, H.S. Jacob and L. T. Furcht. 1985. Am. J. Pathol. 120:13-21.

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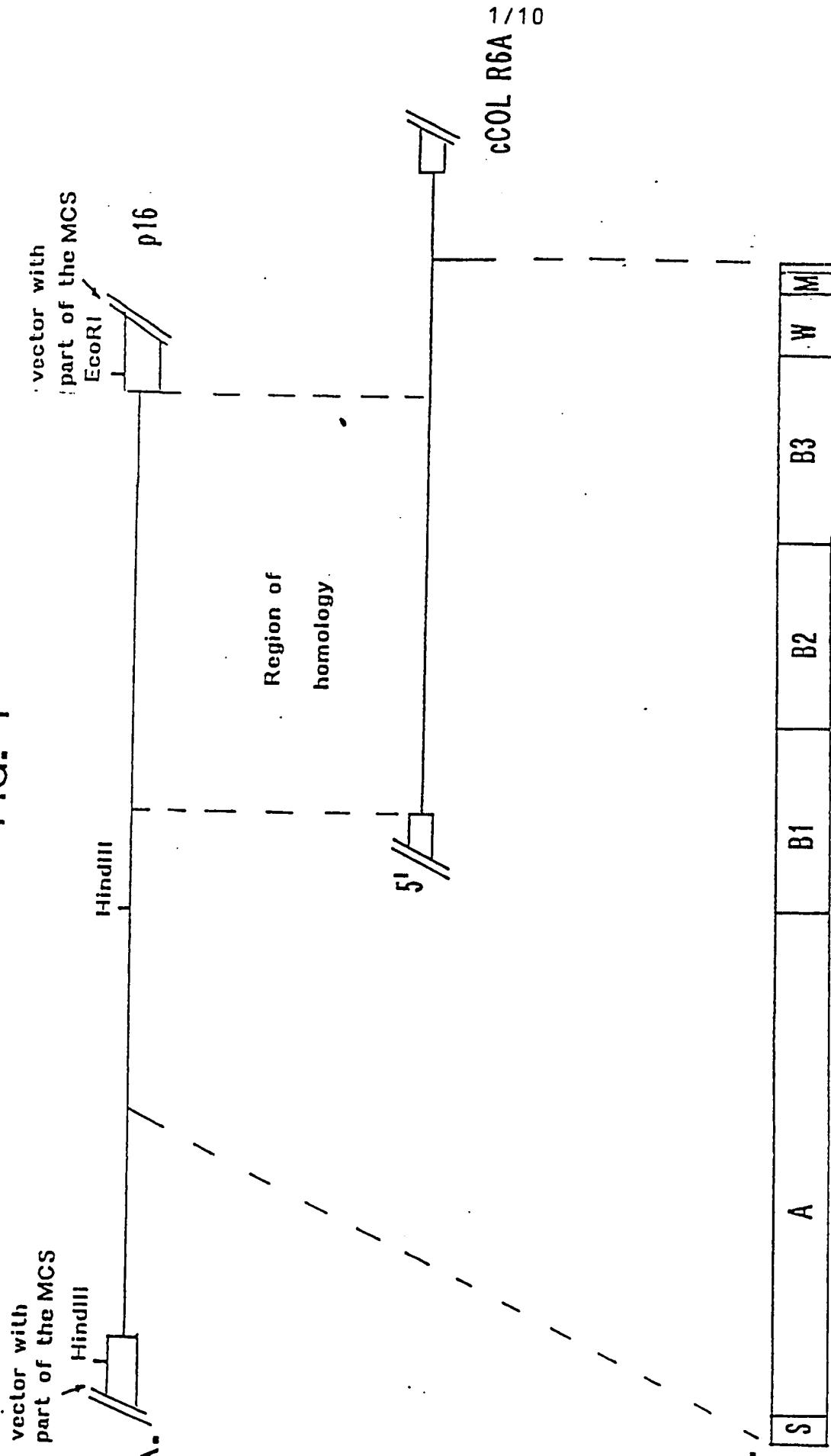
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5 TATAGCTAAT GTAAAATTAA AACTTTCTAA AAAAGATGGA TCAGTTGTAA  
AGGACAATCA AAAAGAAATT GAGATTATAA CAGATGCAA CGGTATTGCT  
AATATTAAAG CGTTGCCTAG TGGAGACTAT ATTTAAAAG AAATAGAGGC  
GCCACGACCG TATACATTG ATAAGGATAA AGAATATCCG TTTACTATGA  
AAGATACAGA TAATCAGGGA TATTTTACGA CTATTGAAAA TGCAAAAGCG  
10 ATAGAAAAAA CAAAAGATGT TTCTGCTCAA AAGGTTGGG AAGGCACCTCA  
AAAAGTGAAA CCAACGATT ATTCAAGTT GTACAAACAA GATGACAATC  
AAAATACAAC ACCAGTAGAC AAAGCAGAGA TTAAAAAATT AGAAGATGGA  
ACGACAAAAG TGACATGGTC TAATCTTCCG GAAAATGACA AAAATGGCAA  
GGCTATTAAT TATTTAGTTA AAGAAGTAAA TGCTCAAGGT GAAGATACAA  
15 CACCAGAAGG ATATACTAAA AAAGAAAATG GTTTAGTGGT TACTAATACT  
GAAAAACCAA TCGAAACAAAC ATCAATTAGT GGTAAAAAAG TATGGGACGA  
CAAAGACAAT CAAGATGGTA AGAGACCAGA AAAAGTCAGT GTGAATTAT  
TGGCTAACGG GGAGAAAAGTA AAAACGTTAG ACGTGACATC TGAAACAAAC  
TGGAAAGTACG AATTAAAGA CTTACCGAAG TATGATGAAG GAAAGAAAAT  
20 AGAATATACA GTGACCGAAG ATCACGTAAA AGACTACACA ACAGACATCA  
ACGGTACGAC AATAACGAAC AAGTATACAC CAGGAGAGAC ATCGGCAACA  
GTAACAAAAA ATTGGGATGA CAATAATAAC CAAGACGGAA AACGACCAAC  
TGAAATCAA GTTGAGTTAT ATCAAGACGG AAAAGCAACA GGAAAAACGG  
CAACATTAAA TGAATCTAAT AACTGGACCC ATACGTGGAC AGGATTAGAT  
25 GAAAAAGCA: AGGACAACA AGTA: ATAC ACAGTCGAGG AATTAACAAA  
GGTCAAAGGT: ATACAACAC ATGTGGATAA CAATGATATG GGTAACTTGA  
TTGTGACGAA TAAATATACG CCAGAAACAA CATCAATTAG TGGTAAAAAA  
GTATGGGACG ACAAAAGACAA TCAAGATGGT AAGAGACCAG AAAAAGTCAG  
TGTGAATTAA TTGGCTGATG GAGAGAAAAGT AAAAACGTTA GACGTGACAT  
30 CTGAAACAAA CTGGAAGTAC GAATTAAAG ACTTACCGAA GTATGATGAA  
GGAAAGAAAA TAGAATATAC AGTGACCGAA GATCACGTAA AAGACTACAC  
AACAGACATC AACGGTACGA CAATAACGAA CAAGTATACA CCAGGAGAGA  
CATCGGCAAC AGTAACAAAA AATTGGGATG ACAATAATAA CCAAGACGGAA  
AAACGACCAA CTGAAATCAA AGTTGAGTTA TATCAAGACG GAAAGCAAC  
35 AGGAAAAACG GCAACATTAA ATGAATCTAA TAACTGGACC CATACTGGAA  
CAGGATTAGA TGAAAAAGCA AAAGGACAAC AAGTAAAATA CACAGTCGAG

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10. Chemical synthesis method for producing a collagen binding protein or polypeptide according to claim 1, whereby an amino acid residue is built up based on said nucleotide sequence encoding for said protein or polypeptide starting 5 from the C-terminal serine, which is stepwise reacted with the appropriate amino acid, whereby it is finally reacted with alanine at the N-terminal end, to form the collagen binding protein or polypeptide.

10 11. A collagen binding protein or polypeptide comprising at least one of the amino acid sequence  
Ala  
ArgAspIleSerSerThrAsnValThrAspLeuThrValSerProSerLysIleGluAsp  
GlyGlyLysThrThrValLysMetThrPheAspAspLysAsnGlyLysIleGlnAsnGly  
15 AspMetIleLysValAlaTrpProThrSerGlyThrValLysIleGluGlyTyrSerLys  
ThrValProLeuThrValLysGlyGluGlnValGlyGlnAlaValIleThrProAspGly  
AlaThrIleThrPheAsnAspLysValGluLysLeuSerAspValSerGlyPheAlaGlu  
PheGluValGlnGlyArgAsnLeuThrGlnThrAsnThrLeuAspAspLysValAlaThr  
IleThrSerGlyAsnLysSerThrAsnValIleGlyTrpIleLysValLysArgGluPro  
20 ValValPheLeuIleAsnLysSerGlyLysIleCysTyrGlnGluAspThrThrHisVal  
ArgTrpPheLeuAsnIleAsnAsnGluLysSerTyrValSerLysAspIleThrIleLys  
AspGlnIleGlnGlyGlyGlnGlnLeuAspLeuSerThrLeuAsnIleAsnValThrGly  
ThrHisSerAsnTyrTyrSerGlyGlnSerAlaIleThrAspPheGluLysAlaPhePro  
GlySerLysIleThrValAspAsnThrLysAsnThrIleAspValThrIleProGlnGly  
25 TyrGlySerTyrAsnSerPheSerIleAsnTyrLysThrLysIleThrAsnGluGlnGln  
LysGluPheValAsnAsnSerGlnAlaTrpTyrGlnGluHisGlyLysGluGluValAsn  
GlyLysSerPheAsnHisThrValHisAsnIleAsnAlaAsnAlaGlyIleGluGlyThr  
ValLysGlyGluLeuLysValLeuLysGlnAspLysAspThrLysAlaProIleAlaAsn  
ValLysPheLysLeuSerLysLysAspGlySerValValLysAspAsnGlnLysGluIle  
30 GluIleIleThrAspAlaAsnGlyIleAlaAsnIleLysAlaLeuProSerGlyAspTyr  
IleLeuLysGluIleGluAlaProArgProTyrThrPheAspLysAspLysGluTyrPro  
PheThrMetLysAspThrAspAsnGlnGlyTyrPheThrThrIleGluAsnAlaLysAla  
IleGluLysThrLysAspValSerAlaGlnLysValTrpGluGlyThrGlnLysValLys  
ProThrIleTyrPheLysLeuTyrLysGlnAspAspAsnGlnAsnThrThrProValAsp  
35 LysAlaGluIleLysLysLeuGluAspGlyThrThrLysValThrTrpSerAsnLeuPro  
GluAsnAspLysAsnGlyLysAlaIleLysTyrLeuValLysGluValAsnAlaGlnGly

FIG. 1



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FIG. 2

(cont'd)

1141	GACATGATTAAAGTGGCATGGCGACAGCGGTACAGTAAAGATAGAGGGTTATAGTAAA AspMetIleLysValAlaTrpProThrSerGlyThrValLysIleGluGlyTyrSerLys	1200
1201	ACAGTACCATTAACGTAAAGGTGAAACAGGTGGGTCAAGCAGTTATTACACCAAGACGGT ThrValProLeuThrValLysGlyGluGlnValGlyGlnAlaValIleThrProAspGly	1260
1261	GCAACAATTACATTCAATGATAAGTAGAAAAATTAAAGTGTGTTGGGATTGCAAGAA AlaThrIleThrPheAsnAspLysValGluLysLeuSerAspValSerGlyPheAlaGlu	1320
1321	TTTGAAGTACAAGGAAGAAATTAAACGCAAAACAAATACTTTAGATGACAAAGTAGCTACG PheGluValGlnGlyArgAsnLeuThrGlnThrAsnThrLeuAspAspLysValAlaThr	1380
1381	ATAACATCTGGGAATAAAATCAACGAATGTTATCGGTTGGATAAAAGTGAAGCGGGAACCA IleThrSerGlyAsnLysSerThrAsnValIleGlyTrpIleLysValLysArgGluPro	1440
1441	GTAGTGTITCTAATTAATAAAAGCGGGAAAGATATGCTACCAAGAAGATACGACACATGTA ValValPheLeuIleAsnLysSerGlyLysIleCysTyrGlnGluAspThrThrHisVal	1500
1501	CGATGGTTTTAAATATTAACAATGAAAAAGTTATGTATCGAAAGATATTACTATAAG ArgTrpPheLeuAsnIleAsnAsnGluLysSerTyrValSerLysAspIleThrIleLys	1560
1561	GATCAGATTCAAGGTGGACAGCAGTTAGATTAAACGACATTAACATTAAATGTGACAGGT AspGlnIleGlnGlyGlnGlnLeuAspSerThrLeuAsnIleAsnValThrGly	1620
1621	ACACATAGCAATTATTATAGTGGACAAAGTGCATTACTGATTGAAAAAGCCTTCCA ThrHisSerAsnTyrTyrSerGlyGlnSerAlaIleThrAspPheGluLysAlaPhePro	1680
1681	GGTTCTAAAATAACTGTTGATAATACGAAGAACACAATTGATGTAACAAATTCCACAGGC GlySerLysIleThrValAspAsnThrLysAsnThrIleAspValThrIleProGlnGly	1740
1741	TATGGGTCAATAATAGTTTCAATTAACTACAAAAACCAAAATTACGAATGAAACAGCAA TyrGlySerTyrAsnSerPheSerIleAsnTyrLysThrLysIleThrAsnGluGlnGln HindIII	1800
1801	AAAGAGTTTGTAAATAATTCAACAGCTGGTATCAAGAGCATGGTAAGGAAGAAGTGAAC LysGluPheValAsnAsnSerGlnAlaTrpTyrGlnGluEisGlyLysGluGluValAsn	1860
1861	GGGAAATCATTTAATCATACTGTGCACAATATTAAATGCTAACGCCGTATTGAAGGTACT GlyLysSerPheAsnEisThrValEisAsnIleAsnAlaAsnAlaGlyIleGluGlyThr	1920
1921	GTAAAAGGTGAATTAAAAGTTTAAACAGGATAAAAGATACCAAGGCTCCTATAGCTAAT ValLysGlyGluLeuLysValLeuLysGlnAspLysAspThrLysAlaProIleAlaAsn	1980
1981	GTAATTAACTTAAACTTCAAAAAAGATGGATCAGTTGTAAAGGACAATCAAAAGAAATT ValLysPheLysLeuSerLysLysAspGlySerValValLysAspAsnGlnLysGluIle	2040
2041	GAGATTATAACAGATGCAACGGTATTGCTAATATTAAAGCGTTGCCTAGTGGAGACTAT GluIleIleThrAspAlaAsnGlyIleAlaAsnIleLysAlaLeuProSerGlyAspTyr ATTTAAAAGAAATAGAGGGGCCACGACCGTATACATTGATAAGGATLALGATTTT	2100

3121	GTATGGGACGACAAGACAAATCAAGATGGTAAAGAGACCAAGAAAAAGTCAGTGTGAATTAA ValTrpAspAspLysAspAsnGlnAspGlyLysArgProGluLysValSerValAsnLeu	3180
3181	TTGGCTGATGGAGAGAAAAAGTAAAAACGTTAGACGTGACATCTGAAACAAACTGGAAAGTAC LeuAlaAspGlyGluLysValLysThrLeuAspValThrSerGluThrAsnTrpLysTyr	3240
3241	GAATTTAAAGACTTACCGAAGTATGATGAAGGAAAGAAATAGAATATAACGTGACCGAA GluPheLysAspLeuProLysTyrAspGluGlyLysLysIleGluTyrThrValThrGlu	3300
3301	GATCACGTAAAAGACTACACAAACAGACATCAACGGTACGACAATAACGAAACAAAGTATAACA AspHisValLysAspTyrThrAspIleAsnGlyThrThrIleThrAsnLysTyrThr	3360
3361	CCAGGAGAGACATCGGCAACAGTAACAAAAAATTGGGATGACAATAATAACCAAGACGGA ProGlyGluThrSerAlaThrValThrLysAsnTrpAspAspAsnAsnGlnAspGly	3420
3421	AAACGACCAACTGAAATCAAAGTTGAGTTATATCAAGACGGAAAAGCAACAGGAAAAACG LysArgProThrGluIleLysValGluLeuTyrGlnAspGlyLysAlaThrGlyLysThr	3480
3481	GCAACATTAATGAAATCTAAATACTGGACCCATACGTGGACAGGATTAGATGAAAAAGCA AlaThrLeuAsnGluSerAsnAsnTrpThrHisThrTrpThrGlyLeuAspGluLysAla	3540
3541	AAAGGACAACAAGTAAAATACACAGTCGAGGAATTAAACAAAGGTCAAGGTATACAAACA LysGlyGlnGlnValLysTyrThrValGluGluLeuThrLysValLysGlyTyrThrThr	3600
3601	CATGTGGATAACAACTGATATGGGCAACTTGATTGTGACGAATAATATAACGCCAGAAACA HisValAspAsnAsnAspMetGlyAsnLeuIleValThrAsnLysTyrThrProGluThr	3660
3661	ACATCAATTAGCGGTGAAAAAGTATGGGACGACAAGACAATCAAGATGGTAAAGACCA ThrSerIleSerGlyGluLysValTrpAspAspLysAspAsnGlnAspGlyLysArgPro	3720
3721	AAAAAAGTCAGTGTAAATTATGGCTAACGGAGAGAAAAAGTAAAAACGTTAGACGTGACA GluLysValSerValAsnLeuLeuAlaAsnGlyGluLysValLysThrLeuAspValThr	3780
3781	TCTGAAACAACTGGAAAGTACGAATTAAAGACTTACCGAAGTATGATGAAGGAAAGAAA SerGluThrAsnTrpLysTyrGluPheLysAspLeuProLysTyrAspGluGlyLysLys	3840
3841	ATAGAATATAACGTGACCGAAGATCACGTAAAAGACTACACAAACAGACATCAACGGTACG IleGluTyrThrValThrGluAspHisValLysAspTyrThrAspIleAsnGlyThr 3' end of insert in p16 ← ACAATAACGAAACAGTATAACCCAGGAGAGACATCGGCAACAGTAACAAAAAATTGGGAT	3900
3901	ThrIleThrAsnLysTyrThrProGlyGluThrSerAlaThrValThrLysAsnTrpAsp	3960
3961	GACAATAATAACCAAGACGGAAAACGACCAACTGAAATCAAAAGTGTGAGTTATCAAGAT	4020

FIG. 2

- (cont'd)

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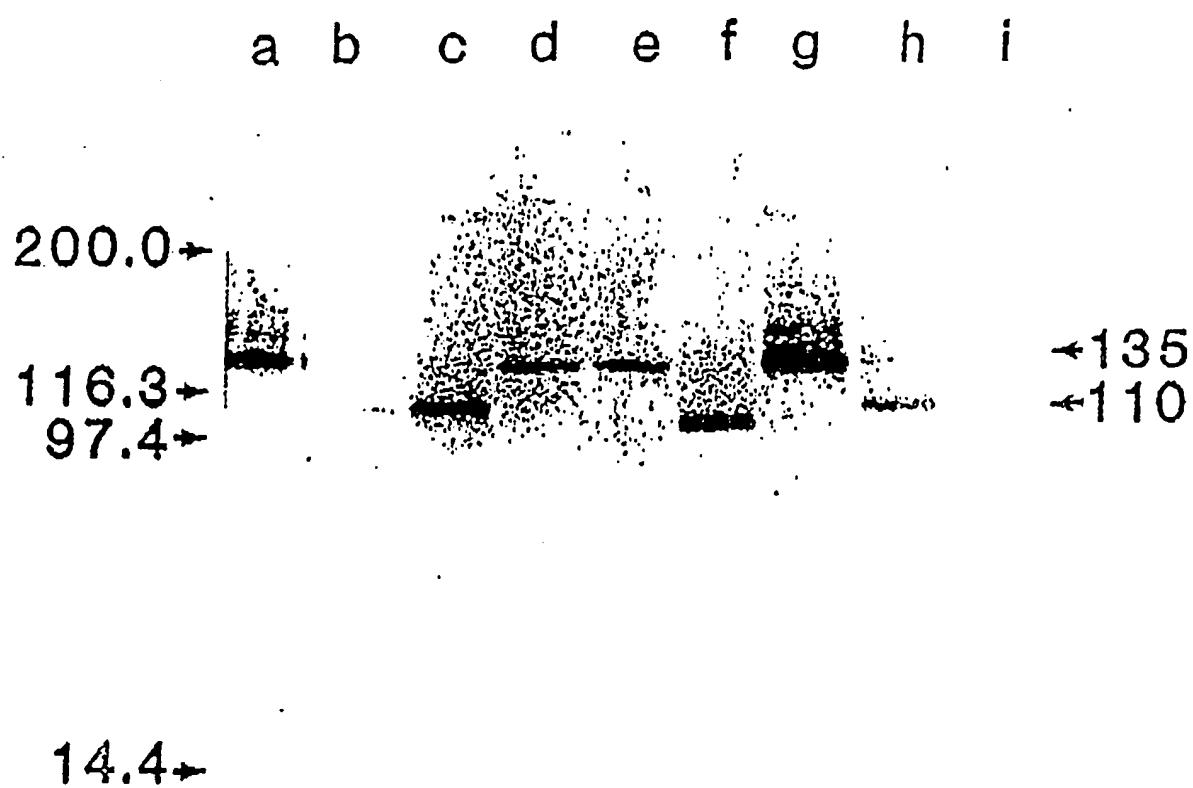


FIG. 3

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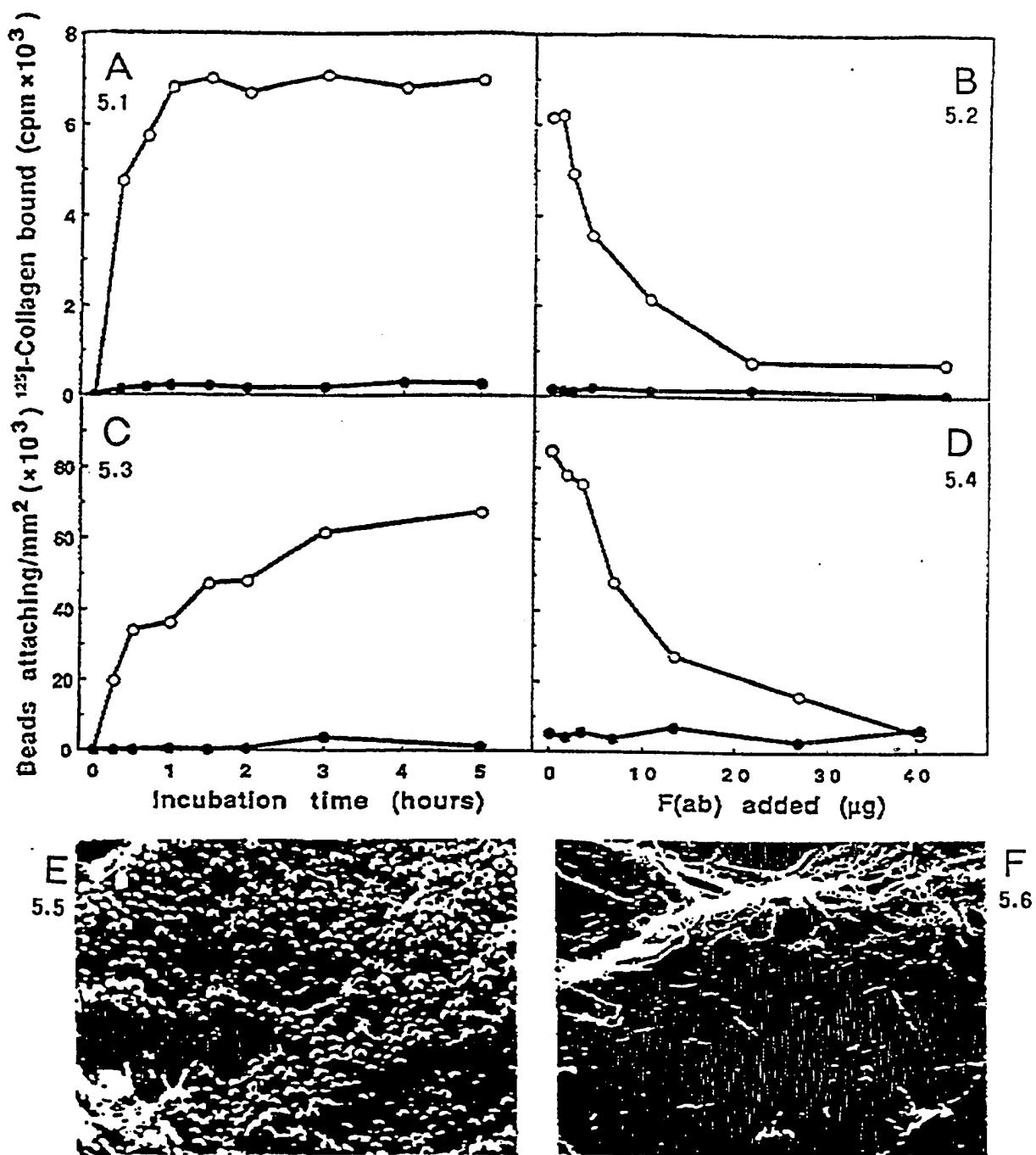
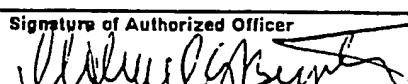


FIG. 5

# INTERNATIONAL SEARCH REPORT

International Application No PCT/SE 91/00707

<b>I. CLASSIFICATION OF SUBJECT MATTER</b> (if several classification symbols apply, indicate all) <sup>6</sup>		
According to International Patent Classification (IPC) or to both National Classification and IPC		
IPC5: C 07 K 15/04		
<b>II. FIELDS SEARCHED</b>		
Minimum Documentation Searched <sup>7</sup>		
Classification System	Classification Symbols	
IPC5	C 07 K; C 12 N	
Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in Fields Searched <sup>8</sup>		
SE,DK,FI,NO classes as above		
<b>III. DOCUMENTS CONSIDERED TO BE RELEVANT<sup>9</sup></b>		
Category	Citation of Document, <sup>11</sup> with indication, where appropriate, of the relevant passages <sup>12</sup>	Relevant to Claim No. <sup>13</sup>
X	EP, A2, 0163623 (ALFA-LAVAL AGRI INTERNATIONAL AB) 4 December 1985, see page 8 lines 7-15 --	1-11
X	Dialog Information Services, file 55: BIOSIS 85-92, Dialog accession no. 7397648, Switalski L M et al: "Isolation and characterization of a putative collagen receptor from staphylococcus- aureus strain cowan 1", & J Biol Chem 264 (35) 1989 21080-21086, -- -----	1-11
<p><b>* Special categories of cited documents:</b><sup>10</sup></p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance, the claimed invention cannot be considered novel or cannot be considered to involve an inventive step</p> <p>"Y" document of particular relevance, the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.</p> <p>"&amp;" document member of the same patent family</p>		
<b>IV. CERTIFICATION</b>		
Date of the Actual Completion of the International Search	Date of Mailing of this International Search Report	
3rd February 1992	1992 -02 - 06	
International Searching Authority	Signature of Authorized Officer	
		

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